RESEARCH ARTICLE

An Improved Synthesis of (*S*)-5-chloro-*N*-{[2-oxo-3-[4-(3-oxomorpholin-4-yl)phenyl]oxazolidin-5-yl]methyl} thiophene-2-carboxamide

SRIDHAR PRASANGI^{1*}, SRINIVAS REDDY DESIREDDY², VELIVELA SRINIVSASA RAO², Y. L. N. MURTHY¹ and MAHESH PALLA¹

¹Department of PNCO, Andhra University, Visakhapatnam-530003, India ²Optimus drugs Ltd., Hyderabad *ridhar.prasangi@gmail.com*

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Abstract: A new and improved process has been developed for the oral anticoagulant rivaroxaban with 99.9% purity and 57.7% overall yield (three main steps). Formation and control of four observed impurities is described.

Keywords: Anticoagulant, Rivaroxaban, Synthesis, Impurity

Introduction

Rivaroxaban (1), chemically (Figure 1) known as 5-chloro-*N*-({(5S)-2-oxo-3-[4-(3-oxomorpholin-4-yl)phenyl]-1,3-oxazolidin-5-yl}methyl)thiophene-2-carboxamide, is a novel, oral, selective direct inhibitor of factor Xa developed by Bayer Healthcare^{1,2}. It has been approved by EMEA and FDA for the prevention of venous thromboembolism in adult patients after total hip replacement or total knee replacement surgery³⁻⁶. Rivaroxaban is available in the market under the brand name Xarelto[®] as 10mg tablets in Europe and US.



Figure 1. Structure of Rivaroxaban (1)

Experimental

The ¹H and ¹³C NMR spectra were recorded in CDCl₃ and DMSO-*d*6 using Brücker 500 MHz NMR spectrometer. The chemical shifts are reported in δ ppm relative to TMS. The LC-MS and mass spectrum was recorded on a Waters XEVO LC-MS spectrometer. The melting points were determined using the capillary method on a YRT-3 melting point

apparatus, which are uncorrected. HPLC analysis was performed on a Shimadazu SPD-15C instrument with a UV detector using Agilent TC-C18 (250 mm × 4.6 mm, 5 μ m) column. Mobile phase A: 0.79 g of ammonium bicarbonate was dissolved in 1000 mL water and added 2.0 mL triethylamine, then adjusted the pH to 8.0 with formic acid. Mobile phase B: acetonitrile. Gradient: 0 min: 80% A, 20% B; 10 min: 80% A, 20% B; 18 min: 68% A, 32% B; 44 min: 34% A, 66% B; 50 min: 34% A, 66% B; 50.1 min: 80% A, 20% B; 60 min: 80% A, 20% B; UV detection at 250 nm; flow rate: 0.7 mL/min; Column oven temperature: 25 °C. The HPLC analysis data is reported in area % and is not adjusted to weight %. 4-(4-aminophenyl)morpholin-3-one (**4**) was provided from Optimus Drugs Ltd.

Ethyl N-[4[(3-oxo-4-morpholinyl)phenyl]carbamate (9)

4-(4-Aminophenyl)morpholin-3-one (1.23 kg, 6.40 mol) and K₂CO₃ (1.07 kg, 7.74 mol) were suspended in 6.75 L toluene and 2.05 L water at 0-5 °C. To the stirred suspension solution of ethyl chloroformate (0.91 kg, 8.39 mol) in 2.7 L toluene was slowly added at the same temperature and continued stirring for 3 h at room temperatue. Subsequently, the mixture was added 6.75 L water and stirred for 30 min. The precipitant was filtered and washed with 1.5 L water. The crude solid was slurried with ethyl acetate (6.75 L) for 2 h. The product was collected by filtration and dried under vacuum to give **9** as a white solid (1.49 kg, 88.1%) with % purity by HPLC. m. p. 190.3-193.1 °C; ¹H NMR (CDCl₃) δ : 1.29 (3 H, m), 3.73 (2 H, m), 4.08 (2 H, m), 4.03 (2 H, m), 4.33 (2 H, s), 6.76 (1 H, s), 7.27 (2 H, d, *J*=9.0 Hz).

(s)-1-Chloro-3-[(4-chloro-E-benzylidene)-amino]-propan-2-ol (10)

Compound **10** was synthesized from (s)-epichlorohydrin according to the reported procedure with minor modification²². To a solution of *p*-chlorobenzaldehyde (1.48 kg, 10.53 mol) in 6.0 L methanol was added 1.03 L aq. ammonia (28 wt%, mol) in one portion, the resulting mixture was stirred for 20 min at room temperature then (*s*)-epichlorohydrin (986.0 g, 15.27 mol) was charged in single portion. The reaction mixture was stirred for 12 h then heated to 40 °C and continued stirring for 2 h. After cooling to room temperature, 9.0 L water was added and methanol was evaporated under vacuum below 45 °C. The precipitant was filtered and slurried with 5.0 L petroleum ether. The solid was filtered and then dissolved in 5.0 L dichlormethane. The organic phase was separated and dried with anhydrous Na₂SO₄. The filtrate containing compound **10** was used directly for the preparation of **3.HCl** without isolation.

4-{4-[(5S)-5-(Aminomethyl)-2-oxo-1,3-oxazolidin-3-yl]phenyl}morpholin-3-one hydrogen chloride (**3 HCl**)

To a suspension of **10** (1.43 kg, 5.41 mol) in 2.25 L CH₂Cl₂, Lithium *t*-butoxide (1.17 kg, 10.42 mol) was added at 5-10 °C in three portions. After being stirred for 30 min at room temperature, the above solution containing compound **10** (~8.11 mol) was added over 30 min. The resulting solution was heated to reflux for 12 h then cooled to room temperature, 16.4 L ethanol was added and the pH of the solution was adjusted to 1.0-2.0 using concentrated HCl (1.6 L, mol). The reaction mixture was stirred for 1.5 h at room temperature, and then the solid was filtered and washed with ethanol (1.6×2 L). The filtrate was stored for the reuse of *p*-chlorobanzaldehyde. The wet solid was charged into ethanol (11.0 L), heated to reflux and stirred for 30 min. The resultant mixture was cooled to 25-30 °C and stirred for 60 min. The product was filtered, washed with ethanol (1.1×2 L) and dried at 45 °C under reduced pressure (400 mmHg) to afford 1.45 Kg (81.8%) of 3 HCl as an off-white solid.

The filtrate containing *p*-chlorobenzaldehyde was added into 25 L water, the organic phase was separated and washed with weter (2.5×2 L). After drying (Na₂SO₄) and evaporation of the solvent, the residue was distilled under reduced pressure to give g *p*-chlorobenzaldehyde (0.77 kg, 92-94 °C/10 torr), which could be reused for the synthesis of compound **10**.

5-Chlorothiophene-2-carbonyl chloride (8)

5-Chlorothiophene-2-carboxylic acid (0.75 kg, 4.61 mol) was suspended in 2.6 L toluene and heated to 80 °C. Thionyl chloride (640 mL, 8.81 mol) was added dropwise over a period of 20 minutes at room temperature; the reaction mixture was heated to 120 °C and stirred for 2 h at the same temperature. The reaction was monitored by TLC (charged one drop of reaction solution in 1 mL methanol) until gas evolution ceases. After removal of the excess thionyl chloride and toluene by evaporation under vacuum, the residue was dissolved in toluene (1.5 L) for the next step.

Rivaroxaban (1)

3'HCl (1.40 kg, 4.27 mol) was dissolved in 4.85 L water and filtered, to the filtrate 6.0 L toluene and Na₂CO₃ (0.56 kg, 5.28 mol) was added at room temperature. The mixture was cooled to 8 to 12 °C, the above solution of **9** in toluene was then added, and the reaction mixture was stirred for 2 h at room temperature. After completion of reaction, 2.5 L acetone was added and the precipitated solid was filtered and washed with 2 mol/L aq. HCl (7.0 L). Wet solid was dried at 70 °C under reduced pressure (400 mmHg) to afford 1.76 kg of the crude product **1** with 99.0% purity by HPLC. The crude product was charged into acetic acid (7.8 L) and heated to reflux for 15 min. The clear solution was cooled to 15 °C and stirred for 2 h and the precipitated solid was filtered and washed with 1.5 L acetone. The wet solid was dried at 70 °C under reduced pressure (400 mmHg) to furnish 1.49 kg (80.0%) of the final product with 99.9% purity by HPLC.

¹H NMR (DMSO) δ: 3.61 (2 H, t, *J*=5.4 Hz), 3.71 (2 H, t, *J*=5.4 Hz), 3.85 (1 H, m), 3.97 (2 H, t, *J*= 4. 5 Hz), 4.19 (3 H, t, *J*=7. 5 Hz), 4.84 (1 H, m), 7.19 (1 H, d, *J*=4.2Hz), 7.40 (2 H, d, *J*=9.0 Hz), 7.57 (2 H, t, *J*=9.0 Hz), 7.69 (1 H, d, *J*= 4.19 Hz), 8.96 (1 H, t, *J*=5.7 Hz).

Many methods have been reported to date for the synthesis of rivaroxaban⁷⁻¹². Most of them share the use of 5-*s*-hydroxymethyl oxazolidione **2** or 5-*s*-aminomethyl oxazolidione **(3)** as key intermediate. However 5-*s*-hydroxymethyl group must be activated as sulfonate then displaced with excess ammonia in sealed vessel¹³ or substituted with potential explosive NaN₃ then hydrogenated^{14,15} to generate the corresponding aminomethyl group of rivaroxaban. So more efforts have been made on the synthesis of 5-*s*-aminomethyl oxazolidione **3** as key intermediate (Figure 2).



Figure 2. Structure of 5-s-hydroxymethyl oxazolidione (2) and 5-s-aminomethyl oxazolidione (3)

The synthesis of rivaroxaban via 4-[4-[5-(Aminomethyl)-2-oxo-1,3-oxazolidin-3-yl] phenyl]-morpholin-3-one was originally reported² in WO0147919, the synthetic route shown in Scheme 1, Initially (R)-2-(2-hydroxy-3-{[4-(3-oxomorpholin-4-yl) phenyl)amino]-1*H*-isoindole-1,3-(2*H*)-dione (**6**) is obtained by reaction between 4-(4-amino-phenyl)morpholin-3-one (**4**) and (s)-2-[2-oxiranylmethyl]-1*H*-isoindole -1,3-(2*H*)-dione (**5**) in an ethanol/water mixture, then compound **6** reacted with *N*, *N*-carbonylimidazole (CDI) in

the presence of a catalytic amount of DMAP using THF as solvent to get compound 2-[[(5S)-2-Oxo-3-[4-(3-oxo-4-morpholinyl) phenyl]-1, 3-oxazolidin-5-yl] methyl]-1Hisoindole-1, 3(2H)-dione (7), then phthalimide protective group of 7 is removed by reaction with aq. CH_3NH_2 to give 5-s-aminomethyl oxazolidione (3), which was acylated with 5chlorothiophene-2-carbonyl chloride (8) using pyridine as solvent and acid scavenger to afford rivaroxaban (1) with HPLC purity of 100% after column chromatography purification. Berwe¹⁶ and Mali¹⁷ et al., disclosed some optimization of reaction conditions described in the WO 0147919 to make this process suitable for scale-up in a commercial plant. Unfortunely the difficulty in reuse of the phthalimide protecting group after hydrazinolysis or aminolysis enable this process less green and cheap. Several other routes have been reported on the synthesis of 5-(s)-aminomethyl oxazolidione (3), however, there are several drawbacks associated with the process described in these reported patents and papers. These drawbacks include the use of tedious chromatography for purification¹⁸, use of potential explosive¹⁹ or inflambe reagent during the reaction²⁰ and low yield of the intermediate and final product²¹. Therefore there is a need for improved process of rivaroxaban for industrial application.



Results and Discussion

The structure of oxazolidinone is a magic medicinal feature in many medicines including linezolide, rivaroxaban and cethromycin. Inspired by the process to linezolide²², a similar protocol was taken to building the oxazolidinone moiety. 4-(4-Aminophenyl)morpholin-3one (4) was reacted with ethyl chloroformate in a mixture of toluene and water using K_2CO_3 as acid scavenger to afford carbamate (9). Carbamate (9) has poor solubility in water and toluene which lead to ready isolation through filtration. As reported that 4-chlorohydrin imine analogue was highly crystalline²³ and choosed in the oxazolidione formation, then the cyclization of carbamate (9) with chlorohydrin imine (10) proceeded in refluxing CH_2Cl_2 in the presence of lithium *t*-butoxide. The reaction could be complete within 12 h without the formation of significant impurities, however, isolation and purification of the desired imine 4-[4-[(5S)-5-[[[(4-Chlorophenyl) methylene] aminol coumpound methyl]-2-oxo-3oxazolidinyl] phenyl]-3-morpholine (11) were problematic as it is prone to cleave to 3 during the work-ups. To overcome this problem, the isolation of imine 11 from the reaction mixture was not carried out immediately when the reaction finished. Reaction mass was acidified to pH 2-3 using a solution of HCl and ethanol to precipitate 3.HCl.

As we expected the aminomethyl oxazolidione HCl salt was precipitated from the reaction mixture which was filtrated and dried for the next step. It was noteworthy to mention that the reaction using benzyl carbamate and methyl carbamate gave the required product with 28% and 75% yields, respectively.



The compound 4-[4-[(5S)-5-(Aminomethyl)-2-oxo-1,3-oxazolidin-3-yl]phenyl]morpholin-3-one HCl (**3.HCl**) was dissolved in water then subjected to acylation with 5chlorothiophene-2-carbonyl chloride (prepared from 5-chlorothiophene-2-carbonylic acid **12**) in toluene using Na₂CO₃ as acid binding agent according to the literature method, rivaroxaban (**1**) was precipitated out with a good yield but failed to eliminate dichloro impurity **A** and deschloro impurity **B** after recrystallization (Figure 3). It is obvious that these two impuritie are derived from the raw material 5-chloro-furan-2-carboxylic acid, in which unreacted and excess chlorination byproducts may be present. To our delight the source of these two impurities could be further controlled in starting material by choosing reliable supplier.



Figure 3. Structure of impurities A & B

The synthetic route shown in Scheme 2 has been used to manufacture several batches of the API rivaroxaban. The HPLC analysis rivaroxaban API indicated the presence of two new impurities at 0.15 and 0.05 area % in several batches of rivaroxaban, identified as 'impurity C' and 'impurity D' (Figure 4). Further work demonstrated that these two impurities were difficult to purge during the final recrystallization and suggested that upstream control would be important. Impurity C and D had been isolated from the mother liquor by silica gel column chromatography, and their structures were confirmed by ¹H NMR, ¹³C NMR and ESI HRMS.





N,*N*'-bis[4-(3-oxo-4-morpholinyl) phenyl]-urea

Figure 4. Structure of impurities C & D

The impurity **C** was believed to arise from related impurities in the intermediate **3.HCl**. It is generally believed that there are small amounts of *p*-chlorobenzaldehyde in chlorohydrin imine **10**, where the carbonyl group could be attacked by lithium salt of 4-aryl- 3-morpholinone in the cyclization step to give 2-(1-hydroxyalkyl)-3-morpholinone (**12**). Similar nucleophilic attack of aldehyde by 3-morpholinone lithium salt²⁴ to form penultimate impurity **C** has been noted in the literature report. After the amidation of **12** with 5-chlorothiophene-2-carbonyl chloride to give the impurity **C** as shown in synthetic scheme **3**, the impurity **C** was found to be present at levels greater than 0.15% in the final product. Unfortunately, removal of *p*-chlorobenzaldehyde from the intermediate by slurry wash or recrystallization proved difficult. To our delight, impurity precursors **12** could be removed efficiently after the crude aminomethyloxazolidione **3.HCl** was slurried with ethanol. Proposed formation of impurity **C** during the synthesis of rivaroxaban.



Scheme 3

The structure of impurity **D** as N,N'-bis[4-(3-oxo-4-morpholinyl)phenyl]-urea was also confirmed by synthesis as shown in scheme **4**, the impurity **D** was easily synthesized from 4-(4-aminophenyl)morpholin-3-one **4** with CDI and matched unambiguously with the fractionated sample of the impurity by ¹H NMR and HPLC retention time.



It is obvious that this impurity is generated during the carbamate formation of **9**. The formation of impurity **D** is possible by the aminolysis of carbamate with 4-(4-aminophenyl)morpholin-3-one under the given conditions. It was found that the use of organic base for example Et_3N and pyridine in CH_2Cl_2 increased the formation of this impurity drastically. When the reaction was carried out in a mixture of toluene and water using inorganic base for example Na_2CO_3 and K_2CO_3 as acid scavenger, the content of this impurity could be minimized to 0.5%. This impurity can not be efficiently eliminated by recrystallization in the carbamation and the next cyclization step due to its high crystallizability. Fortunately, this impurity was almost insoluble in water, so we can remove it by filtration from 3.HCl solution in water before the next amidation step.

Conclusion

We have developed a convenient synthesis of rivaroxaban, three steps from 4-(4-aminophenyl)morpholin-3-one with one additional step to prepare chiral intermediate chlorohydrin 10. Four observed impurities of rivaroxaban were characterized by NMR and MS, the origins of formation impurities A-D during the preparation of rivaroxaban were also mapped out. In addition, the method to lower the concentrations of these impurities to levels accepted by ICH is proposed. All the synthesized products were characterized by IR, NMR and Mass spectroscopic data and their melting points were compared with authentic samples.

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References

- 1. Susanne R, Alexander S, Jens P, Thomas L, Josef P, Karl-Heinz S, Peter Reinemer and Elisabeth Perzborn, *J Med Chem.*, 2005, **48(19)**, 5900-5908; DOI:10.1021/jm050101d
- 2. Alexander S, Thomas L, Jens P, Susanne R and Elisabeth P, 2009, WO 0147919A1, 2001-05-07
- 3. Mann K G, Brummel K and Butenas S, *J Thromb Haemostasis*, 2003, **1**(7), 1504-1514; DOI:10.1046/j.1538-7836.2003.00298.x
- 4. Mueck W, Schwers S and Stampfuss J, *Thromb J.*, 2013, **11**, 10; DOI:10.1186/1477-9560-11-10
- Bauersachs R, Berkowitz S D, BrennerB, Buller H R, Decousus H, Gallus A S, Lensing A W, Misselwitz F, Prins M H, Raskob G E, Segers A, Verhamme P, Wells P, Agnelli G, Bounameaux H, Cohen A, Davidson B L, Piovella F and Schellong S, *New Engl J Med.*, 2010, **363**, 2499–2510; DOI:10.1056/NEJMoa1007903
- 6. Biemond B J, Perzborn E, Friederich P W, Levi M, Buetehorn U and Buller H R, Thromb Haemostasis, 2007, **97(3)**, 471–477; DOI:10.1160/TH06-11-0620
- 7. Thomas C R, HBr bromination, WO2004060887A1(US8106192B2), 2004-07-22.
- 8. Gerdes C, Perzborn E, Pohlmann J, Roehrig S, Straub A, Thomas C R and Tuch A, Schlemmer K H, WO2004101557 (A1), 2004-11-25
- 9. Bodhuri P and Weeratunga G, Patent, WO2010124385A1 2010
- 10. Mohan Rao D, Krishna Reddy P and Venkat Reddy B, PCT WO 2013046211A1, 2013.
- 11. Singh P K, Hashmi M S, Sachdeva Y P and Khanduri C H, Patent, PCT WO 2013156936A1, 2013.
- 12. Kumar A V, Rameshchandra U A, Mansukhlal T N and Kanaksinh V H, Patent, PCT WO 2015026761, 2015.
- 13. Rao D M and Reddy B V, Patent, WO2013105100A1, 2013
- 14. Rafecas Jané L, Comely A C, Ferrali A, Amela Cortés C and Pastó Aguilà M, WO2011080341
- 15. Maroju S, Kumar P N, Maroju R, Velupula G, Haq A and Prasad T R, *J Applicable Chem* (Lumami, India), 2014, **3**(6), 2573-2585
- 16. Berwe M, Thomas C, Rehse J and Grotjohann D, Patent WO2005068456.
- 17. Mali A C, Deshmukh D G, Joshi D R, Lad H D, Patel P I, Medhane V J and Mathad V T, *Sustain Chem Process.*, 2015, **3**(1), 1.
- 18. Yuan J, Liu K, Li L, Yuan Y, Liu X and Li Y, *Molecules*, 2014, **19(9)**, 14999-15004; DOI:10.3390/molecules190914999

- 19. Li C, Liu Y, Zhang Y, and Zhang X. J Chem Res., 2011, 35(7), 400-401; DOI:10.3184/174751911X13098778358582
- 20. Reddy V R, Rao V V S and Reddy D S, Indian patent application IN 3375/CHE/2012.
- 21. Masse, C. E. Masse, Craig. E, Patent, PCT WO2009023233, 2009.
- 22. Imbordino R J, Perrault W R and Reeder M R, Patent, WO 2007116284 A1 20071018
- Perrault W R, Keeler J B, Snyder W C, Clark C L, Reeder M R, Imbordino R J, Anderson R M, Ghazal N, Secreast S L and Pearlman B A, Edited by Koenig S G, Scalable Green Chem., 2013, 157-166; DOI:10.1201/b15466-8
- 24. Dobrev A, Nechev L and Ivanov C, Liebigs Ann Chem., 1989, 8, 815-818.